NATIONAL PROTOCOLS

Programme: Antenatal haemoglobinopathy screening programme

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Foreword

The 2002 report ‘Fair for all: Improving the Health of Ethnic Minority Groups and the Wider Community in Scotland’ and the subsequent guidance letter ‘Fair for all: Working Together Towards Culturally Competent Services’ which was issued later that same year, highlighted the need for more work to be undertaken to meet the specific health needs of the differing ethnic minority communities emerging in Scotland. In response to this the National Resource Centre for Ethnic Minority Health (NRCEMH) was established.

In December 2004, NRCEMH published the outcome of the needs assessment in regard to antenatal and newborn screening policy for sickle cell disorders (SCD) and thalassaemia that it had conducted on behalf of the then Scottish Executive Health Department (SEHD). As part of the needs assessment process the group reviewed two Health Technology Assessments (HTA) on screening for haemoglobinopathies published in 1999 and 2000. The guidance in this report also took account of advice from the National Screening Committee (NSC) Sickle Cell and Thalassaemia Screening Programme and current practice in Scotland, Wales and Northern Ireland.

CEL 31 (2008) issued in July 2008 set out a number of changes to the pregnancy and newborn screening programmes, including the introduction of a standardised programme of screening for sickle cell disorders and thalassaemia in pregnancy and sickle cell disorders in newborn babies. The pregnancy haemoglobinopathy screening programme is based on the low prevalence screening model utilised in England.

Patient Pathway

Pregnancy Screening Pathway for Sickle Cell and Thalassaemia

Screening to occur as early as possible during pregnancy, ideally by 10 weeks but can be done at any time, as soon as a woman arrives for care.

Key
- FOQ = Family Origin Questionnaire
- PND = Prenatal Diagnosis
- SWHMR = Scottish Women's Held Maternity Record

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pregnant

- provide and discuss screening information, give 'your guide to screening tests during pregnancy' leaflet ideally 48 hours in advance of testing

  - offer screening
  - complete FOQ

  - accept screening test

  - bloods and FOQ to haematology laboratory for interpretation

  - offer PND
  - accept PND


Healthcare Professional

- mother - unaffected
- mother - carrier
- mother - affected

  - contact local fetal maternal team as per local policy
  - contact Consultant Obstetrician and then Haematologist, Genetic Counsellor

  - offer option of father testing to mother

  - declines/father unavailable
  - accept
  - offer the father testing

  - father - declines testing
  - father - accepts testing

  - father - unaffected
  - father - carrier
  - father - affected

  - offer referral to Genetic Counsellor & Consultant Haematologist or Fetal Medicine Team (as per local policy)

  - declines
  - accepts
  - discussion and Risk Assessment
  - offer PND


Genetic Counsellor & Obstetric Team

- fetus - unaffected
- fetus - carrier
- fetus - affected

  - counselling & discuss options
  - continue pregnancy
  - termination

- inform obstetric team

- inform GP/GPs of results

- record accept/decline in SWHMR & inform GP

- haematology laboratory process

- molecular genetics laboratory process in liaison with genetics services
1. Introduction

This document contains standard national protocols for all healthcare professionals involved in the NHS Scotland screening programme for haemoglobinopathies in pregnancy. The programme aims to allow informed reproductive choice by identifying couples at risk of an affected pregnancy at an early gestation. In order to ensure equity of service across Scotland, NHS Boards are required to ensure that the screening service provided locally adheres to these protocols.

Standard operating procedures and local protocols are not included in the document; these need to reflect specific local arrangements and therefore need to be produced and maintained locally.

The haemoglobinopathies are a group of inherited blood disorders which affect haemoglobin, the oxygen carrying component of blood. They fall into two main groups—haemoglobin variants (e.g. sickle haemoglobin) which are associated with the production of a structurally abnormal haemoglobin and the thalassaemias, in which there is inadequate production of normal globin chains. Many haemoglobinopathies are of no clinical significance whereas others are associated with severe morbidity and mortality, most notably sickle cell disorders and beta thalassaemia major. Carriers are usually asymptomatic.

Haemoglobinopathies are common in people whose family origins are in malarial parts of the world. In the UK, haemoglobinopathies are seen particularly among minority ethnic groups from Africa, the Caribbean, Southern Europe, South Asia, South East Asia and the Middle East but can be found (less frequently) in all ethnic groups. Approximately 1000 haemoglobin gene variants have been identified worldwide.

Sickle cell disease affects mainly people of African-Caribbean origin and results from the inheritance of sickle haemoglobin (HbS), a variant haemoglobin caused by a point mutation (glu6val) in the beta globin gene. In conditions of low oxygen tension, HbS polymerises within the red cell and the result is a rigid, “sickle” shaped cell. Sickled cells cause vaso-occlusion in the microvasculature and increased fragility of the red cell. The disease is characterised by severe, often painful and life threatening, vaso-occlusive “crises” in the bones, lungs or brain, chronic anaemia, aplastic crises, splenic sequestration, priapism, a risk of overwhelming sepsis due to hyposplenism and chronic organ damage.

Beta thalassaemia is thought to affect more than 700 people, with approximately 214,000 carriers in the UK. The highest prevalence is among Cypriot, Italian, Greek, Indian, Pakistani, Bangladeshi, Chinese, other South East Asian and Middle Eastern populations.

Beta thalassaemia major results in severe transfusion dependent anaemia. Patients require lifelong transfusion support which results in the need for monitoring and therapy to prevent iron accumulation and the resultant associated organ damage.

Both conditions (sickle cell disease and beta thalassaemia major) can restrict a child’s or adult’s ability to conduct normal daily activities, and can also have profound psychosocial effects on individuals and their families.

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2. **Offer of Screening**

The aim of offering screening in pregnancy is to identify couples who are at risk of having an affected child and thereby offer them information on which to base reproductive choices. It is important that screening is offered early to allow time for fathers to be screened so that the results of the screening tests and any prenatal diagnosis (PND) are available sufficiently early for couples to be able to make timely informed choices. There is a known association between gestation at screening offer and uptake of PND, with the early offer of screening being associated with greater uptake of PND.

Screening for sickle cell disorders and thalassaemia should be offered to all women as early as possible in pregnancy, and ideally by 10 weeks. This is in order to offer the full range of reproductive choice, including the offer of termination of pregnancy, if chosen, especially for some groups such as those of Muslim faith; in a way that screening later does not. Screening should be discussed at whatever gestation the woman first presents as this may still be of clinical benefit. It is important that they are aware that this programme differs from the other pregnancy screening programmes, in that it is optimally a two stage screen. For the most accurate analysis of chance, testing of the father of the baby will be requested (irrespective of family origin) if the woman is confirmed as a carrier of or as having a haemoglobinopathy, however screening for that pregnancy using only the woman’s result is also offered if the father does not wish/is not able to be screened, for whatever reason.

All pregnant women will be offered screening for thalassaemia based on a formal process of inspection of routine blood indices. Additionally Scotland will be following the low prevalence screening model for haemoglobinopathies utilised in England and offering women in high risk groups, or women whose partners are in high risk groups, screening for sickle cell disorders and other haemoglobin variants using a recommended Family Origin (Ancestry) Questionnaire (FOQ) to assess risk status (see appendix 1).

### Conditions to be screened for:

| (i) Significant maternal haemoglobinopathies | Hb-SS  
| These should be detected by pregnancy screening and are important for maternal clinical care | Hb-SC  
| Hb-S/D Punjab  
| Hb-SE  
| Hb-SC Arab  
| Hb-S/Lepore and Hb Lepore/ β thalassaemia  
| Hb-S/β thalassaemia  
| Hb-S/δβ thalassaemia  
| Hb H disease (α/-α)  
| β thalassaemia major/intermedia  
| Hb E/β thalassaemia |

| (ii) Maternal conditions requiring partner testing | Hb-SS  
| a) Conditions in (i) | Hb-SC  
| b) Carrier states in mother | Hb-SD Punjab  
| Potential significant disorders in the fetus | Hb-SE  
| Hb-SC Arab  
| Hb-S/Lepore  
| Hb-β/Lepore; Hb-S/β thalassaemia;  
| Hb E/β thalassaemia; β thalassaemia major  
| except cases with silent or near silent |

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<th>HPFH</th>
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c) Any compound heterozygote state including one or more of the above conditions
d) Any homozygous state of the above conditions
2.1 Responsibilities

NHS Scotland is responsible for ensuring that all pregnant women known to the service are provided with clear information in an appropriate format to help them make an informed choice about whether to take up any offer of screening. It is important that the couple are aware that this programme differs from the other pregnancy screening programmes, in that it is optimally a two stage screen. For the most accurate analysis of chance, testing of the father of the baby will be requested if the woman is confirmed as a carrier of or having a haemoglobinopathy, however screening for that pregnancy using only the woman’s result is also offered if the father does not wish/is not able to be screened. Women should be offered advice regarding the advantages and implications of the screen appropriate for their gestation. If the offer of screening is accepted, women (and the fathers undergoing testing) should be made aware of the results of the screen.

NHS Scotland is also responsible for ensuring that all fathers who are offered screening are provided with clear information in an appropriate format to help them make an informed choice about whether to take up any offer of screening. If accepted, it is the maternity services responsibility to arrange for the sample to be taken (either in the maternity setting or arranging for this to be taken elsewhere such as the father’s GP practice). Regardless of where the sample is taken it is the maternity services responsibility to coordinate the sample being taken and being received in the laboratory. Additionally maternity services should ensure that the sample is flagged as a ‘father screen’ being requested as part of the pregnancy screening programme for haemoglobinopathies. The results of the maternal and father sample should be combined to give a risk assessment for that pregnancy and for appropriate counselling to take place.

There are some responsibilities which rest with the woman herself including:
- the registration of pregnancy in time to access pregnancy screening and testing;
- making the decision whether to undergo screening and testing;
- provide accurate clinical information required for the accurate interpretation of the results;
- notifying the NHS if no result is provided within the agreed timeframe;
- attending appointments for onward care, where offered.

Similarly it is the fathers’ responsibility to:
- decide whether to accept screening if offered;
- present for the sample to be taken;
- notify the NHS if no result is provided within the agreed timeframe;
- attend appointments for onward care, where offered.

The father’s result should not be inserted in the maternity notes, however the result of the risk assessment for that pregnancy derived from the combined result of the woman and father and if accepted the PND, should be recorded in the relevant sections of Scottish Woman-Held Maternity Record (SWHMR) and hospital maternity systems. A copy of the father’s result, the outcome of any counselling regarding his results and if required details of any follow on appointments arranged for his onward care, should be issued to his GP.

Each NHS Board should have in place a Multi-Disciplinary Clinical Steering Group to oversee the clinical management, governance and quality of the NHS Boards pregnancy and newborn screening programmes.

The Multi-Disciplinary Steering Group should set out a comprehensive strategic plan for improving quality in accordance with the NHS Board’s overall service developments; develop policies aimed at managing and reducing clinical risk and ensure inter-agency arrangements are in place to support women/couples through the screening and diagnostic pathways.

The Multi-Disciplinary Steering Group also has a responsibility for:
- Contributing to the development and implementation of screening and diagnostic care pathways in line with national standards and policies;
- Ensuring that all care pathways are regularly reviewed and modified in line with the national programme’s changing standards and policy;
- Ensuring arrangements are in place for the audit of the pregnancy and newborn haemoglobinopathy screening programmes and linking to an agreed quality assurance framework;
- Providing a supportive framework for women and their families who have higher risk of, or are found to have a pregnancy at higher chance of a significant haemoglobinopathy;
- Advising and supporting staff on antenatal screening and diagnostic issues;
- Communicating with primary care services;
- Ensuring an ongoing education and training programme is made available for staff offering screening and diagnostic testing to improve awareness and skills and reduce risk of serious untoward incidences;
- Providing an annual screening report which reflects the national minimum audit criteria for the haemoglobinopathy screening programmes.

Quality improvement and performance management are an integral part of any screening programme, to ensure that the programme achieves the highest possible standards. A well developed quality management system should indicate when and how any task is to be undertaken and by whom\(^\text{10}\), with a named person at each step of the pathway. Coherent, consistently applied systems should involve the whole organisation and all the components of the service – from antenatal screening through to the newborn screening and the clinical care of affected children. The whole organisation should be involved in planning, delivering and evaluating the quality of services\(^\text{11}\). This should also be guided by clinical governance processes.

### 2.2 Process

All pregnant women attending an antenatal booking clinic, or being seen in the community, should be given sufficient information on the screening tests available in time to seek more information and make a decision regarding whether to undergo testing. All women and their partners must be given the opportunity to discuss haemoglobinopathy screening options with an appropriately trained professional. Information on how the accuracy of the test can be affected if the pregnancy is the result of either egg or sperm donation should be conveyed sensitively, and if the woman chooses to divulge such information should be counselled appropriately on the clinical information available regarding screening.

All women should be offered haemoglobinopathy screening regardless of their gestation, acknowledging that those being screened later in pregnancy may have fewer management options available than those looking at an earlier gestation. Women, who wish to be counselled regarding chance on their result alone for whatever reason, should be informed that the sensitivity of the tests will be reduced because the father’s information is not available.

If the woman has been tested already, whether in a previous pregnancy or for some other reason, it is recommended by the NSC Sickle Cell and Thalassaemia Programme Centre that the woman should have a routine full blood count taken and MCH and other red cell indices should be reassessed. The recommended policy is that women need not be tested again in the same or a subsequent pregnancy provided that:

- There are two or more previous results from a reputable laboratory, preferably accredited by a body in the UK, which are consistent, unequivocal and well documented
- The red cell indices remain the same and can be used for a reliable interpretation. (If the MCH has been ≥27pg on two previous occasions but the routine blood count in this pregnancy shows the MCH <27pg, the woman need not be tested for thalassaemia).
- The patient identification has three or more matching data items

\(^{10}\) Balmer S. *Quality management for screening: report to the National Screening Committee*: Nuffield Institute for Health

If a previous result is being used this fact must be recorded in the woman’s notes for the current pregnancy. It should be noted that there have been recorded instances where the results do not match when the same person is tested on two different occasions. There could be many reasons for this, from labelling and laboratory errors, to deliberate swapping of identities and duplicate medical record numbers. It may also be due to legitimate technical reasons especially inherent variability of measurements occurring around action values. For this reason there should be local protocols in place that should be followed depending on previous experience and prevalence of such problems. It may be decided to offer retesting in each pregnancy.

NHS Boards should identify a designated person who is responsible for ensuring that:

- Every eligible woman is given the opportunity to be screened;
- Both a primary and failsafe mechanism is in place to ensure that a result is received for all women screened;
- If the woman has a positive screening result partner testing is offered;
- If the woman is in agreement, the baby’s father is given the opportunity to be screened as far as possible (paternity issues should be discussed with sensitivity);
- Both a primary and failsafe mechanism is in place to ensure that a result is received for all fathers screened regardless of whether the sample is taken within the maternity setting or elsewhere;
- Women/fathers have the opportunity to receive the results in writing with the offer of appropriate counselling and onward care into diagnostic pathways.

The host NHS Board of the antenatal clinic/community maternity service is responsible for the clinical governance of the service and for ensuring that:

- Every health professional involved in offering/performing a screening test is suitably qualified and trained;
- Every woman who presents for maternity care is offered screening for haemoglobinopathies;
- Every woman is sensitively made aware of the genetic nature of screening and for accurate results information regarding the biological parents of the fetus would be of relevance (for example if the pregnancy is a result of a donated egg or sperm). Additionally the woman should be made aware of the possibility that the test could reveal the paternity of the pregnancy;
- Every women will be made aware that unless declined they will be screened for thalassaemia through the red blood cell indices. Additionally, from the information provided in the FOQ, the chance of either the women or the father of the baby being a carrier for sickle cell and other haemoglobin variants will be assessed. Further testing will be offered on those assessed to be in the higher chance group;
- It will be made clear that screening will be offered to the partner (father of baby) of all carrier mothers (irrespective of family origin) for sickle cell, other haemoglobin variants and thalassaemia;
- Every woman who books for delivery at hospital or at home is offered screening appropriate to her gestation;
- Information on the offer made and any subsequent father testing offered (and whether or not it is accepted) is recorded in SWHMR or equivalent and relevant maternity systems;
- If screening is accepted the family origin of a woman and her partner should be determined using the FOQ and must be recorded appropriately in SWHMR/relevant maternity systems locally and in the laboratory records;
- Samples are easily identifiable from other haematological specimens for women, and linkage to partner (father) samples must be possible.
3. Consent for Screening

Information should be made available, taking into account of the woman’s physical, cultural, ethical educational and mental health needs, at least 48 hours in advance of the screening tests, unless precluded by late presentation.

Women and their partners should be provided with information about the implications of the screening tests. For haemoglobinopathy screening, this should include:

- implications of receiving high or low risk result;
- information on the false positive rates of the screening test;
- the techniques involved and risks that may be associated with any diagnostic tests and
- information about the conditions themselves.

Screening systems should be discussed as ‘an option’ rather than an inevitable aspect of routine maternity care.

Women and the baby’s father, where applicable, must be given sufficient time to make decisions whenever options are presented.

Where maternal or paternal samples are being obtained, there should be information available about their storage and disposal. Should there be an interest in studying any excess material there should be a process for the woman/father to decline to give consent.

It is the responsibility of the health professional to ensure that the correct information is entered into all fields when completing a screening request card. A FOQ must be submitted with every laboratory request form; even if screening is declined (there is an appropriate field on the FOQ to record this).

If a woman declines a screening or diagnostic test, this should also be recorded in the notes by healthcare professional responsible for her care. A protocol should be in place to allow women who have opted out of screening or diagnostic testing to change their mind and still undergo gestationally appropriate screening or testing at a later date.

Supplementary information, including relevant informative/supportive websites or details of support organisations, should be offered to all women/fathers receiving a positive screening or diagnostic test result.

Professionals involved in screening for haemoglobinopathies should work collaboratively with primary care and appropriate agencies such as social services, voluntary sector support groups, religious bodies and bereavement services in order to provide a comprehensive support network that is centred on the woman’s/father’s needs and requests.
4. Screening for haemoglobinopathies in pregnancy

4.1 Organisational requirements

4.1.1 Family Origin Questionnaire (FOQ)

The questionnaire is aimed at determining the family origins (ancestry) and has been developed by the English programme centre. The classifications are different from the census based self-assigned ethnic group questions. If the FOQ is incorporated into an electronic requesting system, the number of options available must not be fewer than those on the paper version. If any modifications are being considered, for example as part of a combined antenatal screening request form, the programme centre should be contacted for advice.

In Scotland the FOQ should be used as a tool to identify women at high risk and thus determine those who require laboratory screening for haemoglobin variants. The woman should be offered testing if she or her baby’s father is in a high risk group. All women should be screened for thalassaemia using the red cell MCH measurement. The need for testing of the baby’s father in possible cases of $\alpha^0$ thalassaemia will also be determined from the FOQ in conjunction with the red cell indices.

The second use of the FOQ is to assist laboratories with the interpretation of laboratory screening results, particularly in the interpretation of results indicating possible $\alpha$ or $\beta$ thalassaemia. For $\alpha$ thalassaemia, people originating from South East Asia and Southern Mediterranean countries are at higher chance of $\alpha^0$ thalassaemia (which in the homozygous state results in hydrops fetalis). Where a request has been made for antenatal screening but an FOQ has not been supplied it must be assumed that the woman is from a high risk area.

Family origin is also relevant in the interpretation of red blood cell indices and is essential for accurate DNA prenatal diagnosis to ensure that the relevant genotypes are included in the DNA analysis of fetal samples.

4.1.2 Situations requiring particular care

Fertility treatment – Donor Gametes
If the pregnancy has been achieved by the use of a donor egg then the screening results on the woman will not be informative so the baby’s father should always be tested to ensure that this is not a high risk pregnancy. If donor sperm has been used then it may be appropriate to refer back to the fertility clinic for information with regard to whether the donor was screened for being a carrier of a haemoglobinopathy.

Adoption
If either parent has been adopted, the FOQ information may not accurately reflect the true family origins. Such cases should be treated as high risk and have full laboratory screening.

Bone Marrow transplants
In women who have had a bone marrow transplant, the haemoglobin results on her blood specimen will not necessarily indicate the genetic makeup of the fetus. The baby’s father should always be tested to ensure this is not a high risk pregnancy. Caution should be exercised in the interpretation of any haematology results in this instance. If DNA confirmation of the mother’s status is required then pre-transplant DNA or DNA obtained from hair follicles should be used.

4.1.3 Screening for haemoglobin variants

As recommended by the NSC Sickle Cell and Thalassaemia Programme Centre the low prevalence testing algorithm (appendix 2) should be utilised. It should be noted that some women may not be tested if the ethnic group is incorrectly stated on the Family Origin Questionnaire or hidden and the following conditions may be missed using the proposed algorithm:
• ‘silent’ or ‘near silent’ βthalassaemia trait
• βthalassaemia obscured by B12/folate deficiency or liver disease
• possibly some β thalassaemias obscured by severe iron deficiency anaemia
• δβ thalassaemia carriers with HbF< 5% and γδβ thalassaemia carriers
• bthalassaemia occurring outside the defined at risk ethnic groups
• dominant haemoglobinopathies in the partner when the woman is Hb AA, but these are very rare and should be suggested by the family history
• any significant haemoglobin silent on HPLC or IEF
• Hb S, C, Dp, E, OArab in the North-European ethnic group.

Samples and forms should be given a number and the details on the form entered into the screening database. The assigned number should be used to link sample, patient details and analytical results and appear on the final report.

The request form must contain fields which conform to the minimum dataset including:
• Patient identifying details (name, date of birth, Community Health Index (CHI) number)
• Hospital attended or other referral source.
• The date the sample was taken.
• Information on the pregnancy needed to interpret the screening results including a completed FOQ.

Routine measurement of blood indices includes measurements of MCH and MCV. MCH should be used to screen for the risk of thalassaemia. These measurements are usually reported for all routine blood counts.

4.1.4 Partner testing

All genetic tests, including screening, undertaken during pregnancy will always include the possible issue of non paternity. This needs to be considered when offering screening tests, and when inviting the woman’s ‘partner’ or ‘baby’s father’ for testing. The role of the health professional is not to judge the situation but to provide clarity and discretion around genetic screening test results. When a carrier woman is identified during antenatal screening, the issue of non-paternity should be considered when offering screening to the ‘baby’s father’. It should be highlighted to the mother that the correct person to be screened is the ‘baby’s father’ so that the pattern of genetic inheritance in the baby can be correctly assessed. When discussing results the need for discretion is essential.

Partner testing is done by the same haematological testing strategy as for maternal phenotype testing. If the partner has a haemoglobinopathy that can interact with the maternal phenotype as depicted in Appendix 4, then the couple should be counselled. If the parents so choose, fresh blood samples should be sent to a DNA referral laboratory with appropriate consent for molecular analysis in preparation for prenatal diagnosis.

If the father has been tested for sickle cell and thalassaemia previously, the same protocol should be followed as for a previously tested mother. It would be prudent to test for a second time to confirm the previous result and then he need not be tested again provided that:-
• There are two or more previous results from a reputable laboratory, preferably accredited by a body in the UK, which are consistent, unequivocal and well documented
• The red cell indices remain the same and can be used for a reliable interpretation or have previously been normal on two occasions.
• His identification has three or more matching data items
5. Laboratory organisation

5.1 General considerations and requirements for haemoglobinopathy screening laboratories

There must be a named senior member of the laboratory staff at medical consultant or clinical scientist consultant level with expertise in haemoglobinopathy diagnosis, to be responsible for the haemoglobinopathy screening service, with defined lines of accountability for all laboratory aspects of the service.

There must be an agreed standard operating procedure, describing the process of laboratory testing from initial receipt of the specimen until dispatch of the report. The laboratory must adopt the testing algorithm defined by the national screening programme, to determine those pregnancies at risk of sickle cell disease or thalassaemia. This testing algorithm sets out the conditions to be tested for and the analytical methods that can be used.

The laboratory should have links with the newborn laboratory including named contacts and should routinely report on all couples identified as ‘at risk’ along with other relevant antenatal information, to the newborn screening laboratory team. If the laboratory is performing PND or PGD it must be a member of the UK Genetic Testing Network (UK GTN) and comply with the quality criteria laid down by the UK GTN Steering Group.

Every laboratory should follow the principles of good laboratory practice including satisfying themselves that they understand the capabilities and limitations of their chosen technique. The equipment and protocol chosen must fulfil the requirements of the screening programme and demonstrate suitable performance on EQA. There must be a documented risk management policy for the laboratory aspects of the antenatal sickle cell and thalassaemia screening service. This should include a risk assessment which describes the steps in the testing protocols where mistakes could occur and the procedures that have been implemented to minimise the risk of the mistake occurring. Where appropriate this should include a policy for samples sent away for further analysis. It should also include documented procedures for the management and reporting of incidents.

Laboratories must be accredited by an appropriate body e.g. United Kingdom Accreditation Service (UKAS), participate in an accredited external quality assessment scheme e.g. UK National External Quality Assessment Service (NEQAS), and be able to demonstrate satisfactory performance as defined by the criteria specified by the External Quality Assessment (EQA) organisers.

Unlike some other pregnancy screening programmes, there is no minimum specimen throughput specified, although where small numbers affect the timeliness of reporting, centralisation may be an option. The introduction of pathology modernisation may have some impact. It is, however, essential that the NHS Sickle cell and Thalassaemia screening programme algorithms are followed and standards are met, including requirements for data collection. Coordination of pathology services across chemistry and haematology can allow the sharing of HPLC equipment, to allow optimal efficiency of the programme. If this is undertaken, it is essential that HbA2 is analysed by a buffer/column elution programme that clearly separates HbA2 from HbA. If some of the analytical aspects of the programme are conducted across laboratories, there must be a risk assessed protocol for all aspects of the process detailing responsibility for coverage; transfer of selected specimens; receipt of results in a timely manner; appropriate reporting and referral to comply with CPA standard E6. It must also be agreed who will provide KPI data and any data for annual returns.

Laboratories that detect few screen positive cases should link with centres of expertise that can provide diagnostic support for presumed positive cases.
Samples should be easily identifiable from other haematological specimens for women, and linkage to partner (father) samples must be possible. The laboratory must agree to collect a minimum dataset of information for monitoring purposes. The laboratory must participate in audit of the screening service at local and national level and provide an annual report, or the necessary data for the preparation of an annual report.

At least 95% of pregnancy screening reports must be issued to an appropriate healthcare professional within 3 working days of receipt of the specimen. In the absence of universal or standardised maternity information systems, the laboratory information management system will be used as a repository of national aggregated data on the screening programme.

5.2 Laboratory analysis, interpretation and reporting of pregnancy screening results

NHS Scotland has agreed to follow the low prevalence standard operating procedures included in the Sickle Cell and Thalassaemia Handbook for Laboratories issued by the NSC Sickle Cell and Thalassaemia programme centre (Third Edition Oct.2012). For full details of the techniques for the measurement and analysis of samples, the interpretation and reporting of pregnancy screening results and advice on referral of antenatal samples to the DNA referral laboratory, please refer to the Laboratory Handbook. Copies can be downloaded using the following link: http://sct.screening.nhs.uk/cms.php?folder=2493

It is recommended that all laboratories participating in the antenatal haemoglobinopathy screening programme in Scotland familiarise themselves with the contents of this publication and use it as a reference guide.

The guidance contained within the laboratory handbook should contain sufficient information to allow all normal and over 95% of abnormal reports to be reported in a standardised manner, however due to the diversity of haemoglobin variants and thalassaemia syndromes, there will always be some situations that require further tests on different samples, or family studies before a useful clinical diagnosis can be achieved.

If the partner is unavailable for testing or his haemoglobinopathy status is unknown, then counselling based on the woman’s result alone should be provided. The NHS Scotland haemoglobinopathies screening programme supports the woman being offered prenatal diagnosis in this situation if she requests it. Prenatal diagnosis for some genotypes of sickle cell disorders can be undertaken without the partner’s DNA. Similarly, prenatal diagnosis for βthalassaemia can be undertaken without the partner’s DNA, although the diagnosis will not be able to be given with as high a degree of certainty if the partner’s mutation is not known.

5.2.1 Referral of samples to the DNA laboratories for haemoglobinopathy mutation analysis

In some cases, it may not be necessary to send blood samples for DNA analysis (eg. couple when both parents’ results suggest a high risk of α thalassaemia). Detailed guidance can be found in the laboratory handbook.

Blood samples will not be analysed without appropriate patient consent. The blood samples must be accompanied with a DNA laboratory referral form where there is confirmation that the patient has given appropriate consent for DNA to be analysed. The laboratory should be notified prior to sending samples.

It has been agreed that all samples from the NHS Scotland Antenatal haemoglobinopathy screening programme are analysed centrally in Edinburgh at:
Molecular Genetics, David Brock Building, Western General Hospital, Edinburgh EH4 2XU Tel: 0131 5371270 Fax: 0131 5371153
5.3 Training and Education
All those directly involved in the provision of pregnancy screening information or services should have an induction to the programme and must undertake regular updating in line with continuing professional development guidance for their profession.

Additional training for more specific aspects of the programme such as specialist counselling for ‘couples at higher chance’ of an affected pregnancy is required.

5.4 Laboratory reports
The results should be reported using the recommended formats stated in the Sickle Cell and Thalassaemia Handbook for Laboratories. The report issued from the laboratory must contain information which conforms to the minimum data set which includes:

- The patient (name, date of birth, CHI number)
- Hospital or other referral source
- The date the sample was taken as this can be essential if a person has had a recent blood transfusion

Computer generated reports conforming to the agreed minimum dataset should be issued by each laboratory. Over 95% of results or at least an interim report should be available within 3 working days of receipt of the sample by the laboratory.

All reports should be communicated to the referrer and on receipt; the details on the report should be checked. If any information is inaccurate, the laboratory should be contacted as soon as possible with the correct information before the woman is notified. The laboratory should recalculate the risk result and issue an amended report. When reports received from third parties are transcribed into internal laboratory information systems, a full and exact copy of the report must be made. It is not acceptable to summarise or leave out information. The transcribed report should always be checked by an appropriate second person to ensure accuracy of transcription. If laboratories have a system to scan a copy into the laboratory information system this would be a preferred alternative. A report or an exact copy/photocopy should also be included in the patient’s notes and included when referrals are made for counselling or advice.

All women who are identified as having a low risk of a pregnancy affected by a significant haemoglobinopathy should have the opportunity to receive the result in writing and for the report to be filed in SWHMR and hospital maternity systems.

For those women who are identified as having a higher risk, the results should be given priority and faxed, telephoned or securely emailed to the referrer depending on prior arrangement between referrer and screening laboratory. This also applies to the results of fathers who are offered and agree to screening. There should be a robust system in place to ensure that any results arriving at the referral source can be identified immediately on their receipt. This will usually involve a phone call from the laboratory to the referrer indicating that a written report is on its way. Fax or email systems should be in a secure location and only accessible to the relevant staff.

Women/fathers who have undergone screening should have previously indicated how they would like to receive any higher risk result and had their preference documented in SWHMR and/or hospital maternity systems.

The result should be communicated to the patient within 3 working days of it being received and an opportunity to attend for a diagnostic appointment given within a further 2 working days.

Copies of all reports should be provided to the General Practitioner (GP) and to the original referral source if that differs from the unit where antenatal care is being provided.
All reports should be retained in electronic format by each laboratory. It should be possible for information and results relating to individual pregnancies held on the laboratories' screening database to be accessed by telephone enquiry from an identifiable and verifiable source.

All laboratory documentation should be retained for appropriate periods of time and then disposed of as specified in the laboratory Standard Operating Procedure.

5.5 Failsafe

There should be a local system in place to ensure that all samples have been received and that a result has been received and acted upon in a timely manner for every woman/father who has been screened. This should include links made to previous results. All screen positive results must be reviewed regularly in conjunction with the relevant maternity unit(s) to review accountability and responsibility.

All results should be sent by the haematology laboratory to the referral source unless indicated otherwise. If no result has been received within the timeframe agreed with the haematology laboratory, the laboratory should be contacted and should provide a report as soon as possible. If no sample or request was received at the laboratory a repeat sample should be taken and sent to the laboratory as soon as possible.

5.6 The Diagnostic Process

5.6.1 Organisation

It is the responsibility of each NHS Board to ensure that there is a diagnostic pathway for all women/couples identified as having a pregnancy affected by a significant haemoglobinopathy. Women/couples should be counselled by medical and midwifery staff that has specific and recent experience in the diagnostic tests available.

The decision whether to have a diagnostic test should be the woman’s choice. Women should be given time to make that choice even if it involves further appointments. Written information about the diagnostic tests, their techniques and associated risks should be made available at the time the woman is informed that there was a higher chance of a significant haemoglobinopathy from the screening tests. Further information about the conditions themselves with local figures about techniques and number of tests performed by operators (but not necessarily about local miscarriage rates – see Royal College of Obstetricians and Gynaecologists (RCOG) guideline) should be available at the time of the diagnostic appointment. The option to have a diagnostic test should not be dependent on the intention to terminate, if a significant haemoglobinopathy is identified, but information about the techniques and processes involved in termination should be available if requested.

The invasive tests of chorionic villus sampling (CVS) and amniocentesis should be conducted according to the standards laid down in the RCOG guideline: Amniocentesis and Chorionic Villus sampling, Greentop Guideline No 8 201012. The auditable standards of this guideline should be monitored locally but also be available for national audit subject to the NHS Board’s policy on confidentiality.

5.6.2 Diagnostic results

It must be emphasised that the screening programme for haemoglobinopathies in pregnancy is designed to identify most carriers for sickle cell disorders, thalassaemia and related disorders. The screening programme will not identify every couple at risk for every haemoglobinopathy.

6. Evaluation of the antenatal haemoglobinopathy programme

Audit and monitoring of the screening programme should be performance managed at all health service levels (national and local). An overall understanding of the way the programme is operating will combine reviews of data at a National level for items such as outcome of positive case, training and education, performance of individual laboratories and communication issues with local level information and monitoring on issues of coverage, timeliness of process, care pathways and failsafe arrangements and their adequacy.

All screening programmes are expected to have the appropriate tools to support the minimum criteria for the audit process. This must include clerical support, information technology (IT) equipment/software and networks that link with appropriate data collection systems within the NHS Board.

All abnormal findings subsequently proved to be normal should be kept on a database for the purposes of quality control; confirmed diagnosis should be recorded on/in the:

i. NHS Board’s clinical information system
ii. Woman’s maternity hand held notes (SWHMR)
iii. Woman’s hospital notes

A high standard of cytogenetic and perinatal pathology with feedback to the laboratory departments are an essential element for a screening service.

6.1 Quality control

Laboratory services must be able to provide (as a minimum) from the proportion of the pregnant population that had screening,

- Detection rate (DR):
- Screen positive rate (SPR)

Each NHS Board should aim to regularly carry out an exploratory survey of user views and experiences.
7. Adverse Incidents

As with any screening programme, there is potential for significant adverse incidents. All adverse incidents should be managed appropriately to minimise the risks to, and effects on the patient and participating NHS Boards. NSD’s Pregnancy and Newborn Screening Programme Escalation Procedures13 should be consulted.

An adverse incident can be any of the following:

**Administrative**
- Failsafe procedures not instigated
- Woman/ GP not notified of result

**Laboratory**
- Assay errors
- Interpretation errors
- Failure to analyse sample

**Clinical**
- Misdiagnosis
- Long waiting times through process from positive screening test to confirmed diagnosis

7.1 Procedure

Any healthcare professional involved in the NHS Scotland haemoglobinopathies pregnancy screening programme who becomes aware of a suspected problem should follow agreed NHS Board clinical governance procedures.

Local clinical governance procedures may vary from one NHS Board to another but commonly involve an initial period of local investigation and establishment of extent of the problem followed by external independent peer review, when appropriate.

In all cases associated with the screening programme, there will be a thorough investigation and National Services Division (NSD) will be notified early in the process – at the time of internal investigation. In view of the sensitivities of national screening programmes and the public interest in them, NSD may require an external peer review even if local NHS Board management decide not to invoke this.

If necessary NSD and the NHS Board will meet to discuss and agree what action, if any, is required.

NSD will notify Scottish Government Health and Social Care directorates (SGHSC) and decide if action is needed in other NHS Board areas.

**Note:**
These protocols are to be used in addition to, and do not replace, the Boards’ Clinical / Adverse Incident Reporting Procedures.

8. Confidentiality

Professional staff involved in the screening programme should comply with the provisions of the Caldicott Report. In particular, patient-identifiable information will only be used in clearly defined and monitored circumstances, only when absolutely necessary and should entail the use of the minimum necessary patient-identifiable information.

Access to patient identifiable information should be on a strict need to know basis. Everyone in the organisation should be aware of their responsibilities with respect to patient confidentiality and the organisation should ensure that its use of patient-identifiable information is lawful.

National Services Scotland (ISD and NSD) does not require aggregated information returns on the performance of the screening programme to include patient-identifiable information. Information on clinical activity for national data sets and monitoring must be submitted in anonymised format.
### Appendix 1 - FOQ

#### Screening for Haemoglobinopathies

**Family Origin Questionnaire (FOQ)**

<table>
<thead>
<tr>
<th>Hospital Name</th>
<th></th>
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</thead>
<tbody>
<tr>
<td>Chil No.</td>
<td></td>
</tr>
<tr>
<td>Estimated Delivery Date</td>
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</tr>
<tr>
<td>Surname</td>
<td></td>
</tr>
<tr>
<td>Forename</td>
<td></td>
</tr>
<tr>
<td>Date of Birth</td>
<td></td>
</tr>
<tr>
<td>Address 1</td>
<td></td>
</tr>
<tr>
<td>Address 2</td>
<td></td>
</tr>
<tr>
<td>Postcode</td>
<td></td>
</tr>
</tbody>
</table>

**What are your family origins?**

Please tick all boxes in ALL sections that apply to the woman and the baby's father.

<table>
<thead>
<tr>
<th>A. AFRICAN OR AFRICAN CARIBBEAN (BLACK)</th>
<th>Woman</th>
<th>Baby's father</th>
</tr>
</thead>
<tbody>
<tr>
<td>1/ Caribbean Islands</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2/ Africa (excluding North Africa)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3/ Any other African or African-Caribbean family origins (please write in...)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>B. SOUTH ASIAN (ASIAN)</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>1/ India or African-Indian</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2/ Pakistan</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3/ Bangladesh</td>
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</tbody>
</table>

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<thead>
<tr>
<th>C. SOUTH EAST ASIAN (ASIAN)</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>1/ China including Hong Kong, Taiwan, Singapore</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2/ Thailand, Indonesia, Burma</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3/ Malaysia, Vietnam, Philippines, Cambodia, Laos</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4/ Any other Asian family origins (e.g. Caribbean-Asian) (please write in...)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>D. OTHER NON-EUROPEAN (OTHER)</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>1/ North Africa, South America etc</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2/ Middle East (Saudi Arabia, Iran etc)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3/ Any other Non-European family origins (please write in...)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>E. SOUTHERN &amp; OTHER EUROPEAN (WHITE)</th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>1/ Sardinia</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2/ Greece, Turkey, Cyprus</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3/ Italy, Portugal, Spain</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4/ Any other Mediterranean country</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5/ Albania, Czech Republic, Poland, Romania, Russia etc</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>F. UNITED KINGDOM (WHITE) refer to guidance at the back</th>
<th></th>
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</thead>
<tbody>
<tr>
<td>1/ England, Scotland, N Ireland, Wales</td>
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</table>

<table>
<thead>
<tr>
<th>G. NORTHERN EUROPEAN (WHITE) refer to guidance at the back</th>
<th></th>
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</tr>
</thead>
<tbody>
<tr>
<td>1/ Austria, Belgium, Ireland, France, Germany, Netherlands</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2/ Scandinavia, Switzerland etc</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3/ Any other European family origins, refer to chart (e.g. Australia, N America, S Africa) (please write in...)</td>
<td></td>
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</tbody>
</table>

*Hb Variant Screening Requested by F and/or G (re. request from low risk group)

# Higher risk for alpha zero thalassaemia

<table>
<thead>
<tr>
<th>H. DON'T KNOW (incl. pregnancies with donor egg/sperm)</th>
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<tr>
<th>I. DECLINED TO ANSWER</th>
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<tr>
<th>J. ESTIMATED DELIVERY DATE (please write in if not above)</th>
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<tr>
<th>K. GESTATION AT TIME OF TEST</th>
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</table>

OFFER haemoglobin variant screening to all women if they or their baby's father have answers in a shaded box.

Signed ___________________________  Print Name ___________________________

Job Title ___________________________  Contact Tel No ___________________________  Date ___________________________

(By Health Care Professional completing the form)
Guidance for Health Care Professionals

Screening and Diagnostic Uses of the Family Origin Questionnaire

The Family Origin Questionnaire (FOQ) is principally used as a tool to identify women who are at highest risk of being a carrier or having a baby with a haemoglobin variant or disorder.

The FOQ is also used as a tool by laboratory staff to help with the interpretation of results, particularly in the interpretation of results indicating possible alpha or beta thalassaemia. The family origin is also relevant in the interpretation of red blood cell indices and essential for accurate prenatal diagnosis. More information about its use can be found in the laboratory handbook http://sct.screening.nhs.uk/publications

Therefore you need to ask for the family origins of both the woman AND the baby’s father going back at least 2 generations (or more if possible).

Women with Sickle Cell Disease

Screening will also identify women with sickle cell disease, who should be considered “high risk” requiring specialist care during pregnancy from an Obstetrician and Haematologist, and who should be booked for a hospital delivery.

“Low risk” Family Origins

People with family origins from the countries listed below are considered at low risk for haemoglobin variants.

United Kingdom (White)
England, Scotland, Northern Ireland, Wales.

Northern European (White)
Austria, Belgium, Denmark, Greenland, Iceland, Ireland (Eire), Finland, France, Germany, Luxembourg, Netherlands, Norway, Sweden, Switzerland.

Some populations of the following countries have Northern European origin (countries listed above) and are also at low risk for haemoglobin variants:

Northern European Origin (White)
Australia, North America (USA, Canada), South Africa, New Zealand.
Appendix 2 – Testing algorithm for use in Scotland

* Refer analytical results to consultant for an opinion on the need for a clinical referral or consult the laboratory support service helpline.
** High risk if any ethnic/family origins in China (including Hong Kong), Taiwan, Thailand, Cambodia, Laos, Vietnam, Burma, Malaysia, Singapore, Indonesia, Philippines, Cyprus, Greece, Sardinia, Turkey, or if ethnic/family origin uncertain/unknown.
*** Low risk or high risk as determined by the family origin questionnaire. Note - if baby's father is in high risk ethnic group, test the mother's sample regardless of her family origins.
# In all cases consider coexisting αβthalassaemia if both parents are from a high risk area and MCH <27pg.

Reconsider low risk couples if fetal anaemia/hydrops seen on ultrasound scanning or if family history of hydrops fetalis.
### Appendix 3 - Table of parental carrier state combinations

Table of parental carrier state combinations that give rise to the risk of a fetus with significant sickle cell disease of β-thalassaemia

(Table base on the work of Prof. B. Modell)

<table>
<thead>
<tr>
<th>Carrier of</th>
<th>Hb S</th>
<th>β thal</th>
<th>δβ thal</th>
<th>Hb Lepore</th>
<th>Hb E</th>
<th>Hb O\text{Arab}</th>
<th>Hb C</th>
<th>Hb D\text{Punjab}</th>
<th>HPFH</th>
<th>Not identified as a carrier</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hb S</td>
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<td>Hb Lepore</td>
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<td>Hb E</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Hb O\text{Arab}</td>
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<td>Hb C</td>
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<td></td>
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<tr>
<td>Hb D\text{Punjab}</td>
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<tr>
<td>HPFH</td>
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<tr>
<td>Not identified as a carrier</td>
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**Key:**
- **Red:** Serious risk - refer couple for counselling - prenatal diagnosis to be offered
- **Light Red:** Less serious risk - refer couple for counselling - further investigation may be required
- **White:** Minimal risk
Appendix 4 - HIS Haemoglobinopathy Quality Indicators

Indicator 4.1: Screening for haemoglobinopathies – uptake

Indicator 4.1 measures the delivery of screening for haemoglobinopathies to an eligible population.

4.1 Proportion of pregnant women who are screened for haemoglobinopathies and for whom a conclusive screening result is available

Rationale

One of the objectives of antenatal screening for haemoglobinopathies is to ensure that all pregnant women accepting the offer of screening are actually screened. Information on screening uptake is an essential component in identifying trends and monitoring the effectiveness of service improvement. This is a measure of the delivery of screening to an eligible population. Low uptake may indicate that:

- not all pregnant women have been offered screening
- those offered screening are not accepting screening
- those accepting the screen are not being screened, and/or
- those being screened did not have screening recorded in their notes.

How to measure this indicator

<table>
<thead>
<tr>
<th>Indicator 4.1</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Numerator</td>
<td>The number of pregnant women attending for first antenatal appointment during the reporting period for whom a conclusive screening result is available for haemoglobinopathies</td>
</tr>
<tr>
<td>Denominator</td>
<td>The number of pregnant women attending for first antenatal appointment during the reporting period</td>
</tr>
<tr>
<td>Data sources</td>
<td>Maternity services/haematology laboratory</td>
</tr>
<tr>
<td>Reporting period</td>
<td>April 2013 – March 2014</td>
</tr>
</tbody>
</table>

Note: If the woman has been tested already, whether in a previous pregnancy or for another reason, it is recommended by the National Screening Committee Sickle Cell Disease and Thalassaemia Programme Centre that the woman does not need to be tested again in the same or a subsequent pregnancy provided that:

- there are two or more previous results from a reputable laboratory, preferably accredited by a body in the UK, which are consistent, unequivocal and well documented
- the red cell indices remain the same, and
- the patient identification has three or more matching data items.
Indicator 4.2: Screening for haemoglobinopathies – timeliness

Indicator 4.2 measures the timeliness of screening for haemoglobinopathies.

4.2 Proportion of pregnant women having haemoglobinopathy screening for whom a conclusive screening result is available by 10 weeks’ gestation

Rationale

One of the main objectives of antenatal screening for haemoglobinopathies is to ensure that all pregnant women who accept the offer of screening are tested in a timely manner as identified in National Institute for Health and Care Excellence (NICE) guidelines on antenatal care.\(^1\) The majority of prenatal diagnostic testing currently takes place after 11\(^{th}\) weeks’ gestation, which may be too late to allow parents to make informed and timely reproductive choices. A high proportion of women tested after 10 weeks’ gestation may indicate that:

- screening is being offered outside the specified timeframe
- there is a delay between the screening encounter and availability of results, and/or
- women are presenting later than 10 weeks’ gestation for screening.\(^2,3\)

How to measure this indicator

<table>
<thead>
<tr>
<th>Indicator 4.2</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Numerator</strong></td>
<td>The number of pregnant women attending for first antenatal appointment during the reporting period for whom a haemoglobinopathy screening sample is received by 10 weeks’ gestation</td>
</tr>
<tr>
<td><strong>Denominator</strong></td>
<td>The number of pregnant women for whom a haemoglobinopathy screening sample is received at laboratory during the reporting period</td>
</tr>
<tr>
<td><strong>Data sources</strong></td>
<td>Maternity services/haematology laboratory</td>
</tr>
<tr>
<td><strong>Reporting period</strong></td>
<td>April 2014 – March 2015</td>
</tr>
</tbody>
</table>
Indicator 5: Screening for haemoglobinopathies – family origin questionnaire

Indicator 5 measures whether or not haemoglobinopathy screening is supported by a family origin questionnaire (FOQ).

5 Proportion of haemoglobinopathy samples submitted to the laboratory that are supported by a completed FOQ

Rationale

The FOQ facilitates accurate detection of affected pregnancies, one of the main objectives of the haemoglobinopathy screening programme. Low FOQ completion may result in a higher proportion of affected cases being missed.

How to measure this indicator

<table>
<thead>
<tr>
<th>Indicator 5</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Numerator</strong></td>
</tr>
<tr>
<td>The number of blood samples received for haemoglobinopathy screening during the reporting period that are supported by a completed FOQ</td>
</tr>
<tr>
<td><strong>Denominator</strong></td>
</tr>
<tr>
<td>The number of screening samples received by the laboratory for haemoglobinopathy screening during the reporting period</td>
</tr>
<tr>
<td><strong>Data sources</strong></td>
</tr>
<tr>
<td>Maternity services/haematology laboratory</td>
</tr>
<tr>
<td><strong>Reporting period</strong></td>
</tr>
<tr>
<td>April 2014 – March 2015</td>
</tr>
</tbody>
</table>

Note: A completed FOQ is one where all mandatory fields have been filled in.
**Indicator 6: Screening for haemoglobinopathies – coverage**

Indicator 6 measures whether or not haemoglobinopathy screening has been offered to the biological father of the baby where the need has been identified.

6. Proportion of biological fathers who are offered haemoglobinopathy screening following identification of a pregnant haemoglobinopathy carrier

**Rationale**

The national screening protocol states that one of the aims of haemoglobinopathy screening is to allow informed reproductive choice by identifying couples at risk of an affected pregnancy. This is a two-stage screening process. The first stage is to identify a woman carrier. If a woman carrier is identified, the second stage is to offer testing to the biological father. If testing is not offered to the biological father, the chance of having an affected child may range from 1 in 40 to 1 in 800, depending on the gene variant and ancestry population frequency. This chance is reduced to around 1 in 4 or no chance at all if the father is tested, thereby preventing unnecessary intervention and possible fetal loss.¹⁴

**How to measure this indicator**

<table>
<thead>
<tr>
<th>Indicator 6</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Numerator</strong></td>
<td>The number of biological fathers offered haemoglobinopathy screening where the pregnant woman is found to be a haemoglobinopathy carrier during the reporting period</td>
</tr>
<tr>
<td><strong>Denominator</strong></td>
<td>The number of pregnant women found to be haemoglobinopathy carriers during the reporting period</td>
</tr>
<tr>
<td><strong>Data sources</strong></td>
<td>Maternity services/haematology laboratory</td>
</tr>
<tr>
<td><strong>Reporting period</strong></td>
<td>April 2014 – March 2015</td>
</tr>
</tbody>
</table>

**Note:** The offer of testing to the biological father of the baby requires co-ordination of testing between two healthcare professionals involved in the care of the baby and the biological parents (for example, midwife and GP), while respecting the confidentiality of the biological father. The mention of the partner’s name on the request form to the laboratory would facilitate the identification of 'high chance' couples.
<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Full Form</th>
</tr>
</thead>
<tbody>
<tr>
<td>BSH</td>
<td>British Society of Haematology</td>
</tr>
<tr>
<td>CEL</td>
<td>Chief Executive Letter</td>
</tr>
<tr>
<td>CHI</td>
<td>Community Health Index</td>
</tr>
<tr>
<td>CI</td>
<td>Confidence Interval</td>
</tr>
<tr>
<td>CPA UK (Ltd)</td>
<td>Clinical Pathology Accreditation United Kingdom Limited</td>
</tr>
<tr>
<td>CPD</td>
<td>Continuous Professional Development</td>
</tr>
<tr>
<td>DR</td>
<td>Detection Rate</td>
</tr>
<tr>
<td>EDD</td>
<td>Estimated Date of Delivery</td>
</tr>
<tr>
<td>EU</td>
<td>European Union</td>
</tr>
<tr>
<td>FOQ</td>
<td>Family Origin (Ancestry) Questionnaire</td>
</tr>
<tr>
<td>GMC</td>
<td>General Medical Council</td>
</tr>
<tr>
<td>GP</td>
<td>General Practitioner</td>
</tr>
<tr>
<td>HBO</td>
<td>Haemoglobin variant</td>
</tr>
<tr>
<td>HDL</td>
<td>Health Department Letter</td>
</tr>
<tr>
<td>HIE</td>
<td>Higher Institute of Education</td>
</tr>
<tr>
<td>HPC</td>
<td>Health Professional Council</td>
</tr>
<tr>
<td>HTA</td>
<td>Health Technology Assessment</td>
</tr>
<tr>
<td>IT</td>
<td>Information Technology</td>
</tr>
<tr>
<td>KPI</td>
<td>Key Performance Indicator</td>
</tr>
<tr>
<td>MoM</td>
<td>Multiples of the Median</td>
</tr>
<tr>
<td>NHS QIS</td>
<td>National Health Service Quality Improvement Scotland</td>
</tr>
<tr>
<td>NICE</td>
<td>National Institute for Clinical Excellence</td>
</tr>
<tr>
<td>NMC</td>
<td>Nursing and Midwifery Council</td>
</tr>
<tr>
<td>NSD</td>
<td>National Services Division</td>
</tr>
<tr>
<td>NSS</td>
<td>National Services Scotland</td>
</tr>
<tr>
<td>PND</td>
<td>Prenatal Diagnosis</td>
</tr>
<tr>
<td>QC</td>
<td>Quality Control</td>
</tr>
<tr>
<td>QF PCR</td>
<td>Quantitative Fluorescent-Polymerase Chain Reaction</td>
</tr>
<tr>
<td>RCOG</td>
<td>Royal College of Obstetricians and Gynaecologists</td>
</tr>
<tr>
<td>SGHD</td>
<td>Scottish Government Health Directorates</td>
</tr>
<tr>
<td>SPR</td>
<td>Screen Positive Rate</td>
</tr>
<tr>
<td>SWHMR</td>
<td>Scottish Woman Held Maternity Record</td>
</tr>
<tr>
<td>UK GTN</td>
<td>UK Genetic Testing Network</td>
</tr>
<tr>
<td>UK NEQAS</td>
<td>United Kingdom National External Quality Assessment Service</td>
</tr>
<tr>
<td>UK NSC</td>
<td>United Kingdom, National Screening Committee</td>
</tr>
</tbody>
</table>
## Glossary

<table>
<thead>
<tr>
<th>Term</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Affected pregnancies</strong></td>
<td>Pregnancies in which the fetus has the target condition.</td>
</tr>
<tr>
<td><strong>Alpha thalassaemia major (haemoglobin Barts hydrops fetalis)</strong></td>
<td>A severe anaemia that affects the fetus. No normal fetal haemoglobin is produced and this leads to stillbirth or neonatal death.</td>
</tr>
<tr>
<td><strong>Amniocentesis</strong></td>
<td>An invasive procedure undertaken from about 15 completed weeks (15+0) onwards to obtain a sample of amniotic fluid surrounding the fetus. A needle is passed through the mother's abdomen into the uterus, under continuous ultrasound guidance, and a sample of fluid is withdrawn. The fluid, and cells within it, can be tested for certain conditions such as Down’s syndrome and other chromosomal and inherited disorders. Out of 100 women who have this test from 15 weeks it is likely that one will miscarry as a direct consequence of the procedure.</td>
</tr>
<tr>
<td><strong>Amniotic fluid</strong></td>
<td>The fluid surrounding the fetus in the uterus, which protects it during pregnancy and labour. It contains substances and cells from the fetus, which can be removed by amniocentesis and examined.</td>
</tr>
<tr>
<td><strong>Anomaly</strong></td>
<td>An aberration or change often used related to a gene that may or may not result in a disease or condition.</td>
</tr>
<tr>
<td><strong>Antenatal</strong></td>
<td>The period from conception to birth.</td>
</tr>
<tr>
<td><strong>Autosomal inheritance</strong></td>
<td>Mode of inheritance that is independent of the sex chromosomes. It can be dominant or recessive.</td>
</tr>
<tr>
<td><strong>Autosome</strong></td>
<td>A chromosome not involved in sex determination. The human genome consists of 46 chromosomes – 22 pairs of autosomes and one pair of sex chromosomes (the X and Y chromosomes).</td>
</tr>
<tr>
<td><strong>Beta thalassaemia major</strong></td>
<td>A severe anaemia caused by inheritance of two beta thalassaemia genes, resulting in a lack of normal haemoglobin production. Treatment by regular monthly blood transfusions and drugs to remove excess iron leads to long-term survival. Some affected children can be ‘cured’ by bone marrow transplantation.</td>
</tr>
<tr>
<td><strong>Carrier</strong></td>
<td>An individual who carries a single altered gene for a condition where two altered genes are required for an individual to be affected. The carrier can pass on the gene to their offspring, who may be affected if they also inherit an altered gene from their other parent. A carrier is a heterozygote for the gene carried.</td>
</tr>
<tr>
<td><strong>Carrier testing</strong></td>
<td>Testing to find out if a person who does not show symptoms of a condition nevertheless ‘carries’ a copy of an altered gene which could be passed to his or her children.</td>
</tr>
<tr>
<td><strong>Chorionic villus sampling (CVS)</strong></td>
<td>An invasive procedure performed under ultrasound guidance after 10 completed weeks of pregnancy to obtain a sample of placental tissue, which is taken through either the cervix or the abdomen. The range of chromosomal and genetic conditions that can be detected is similar to those for amniocentesis except that Neural Tube Defects cannot be diagnosed. For every 100 women who have this test from the 11th week in pregnancy one or two will miscarry.</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>Chromosome anomaly</td>
<td>A change in the number or arrangement of the normal 23 pairs of chromosomes.</td>
</tr>
<tr>
<td>Confirmed result</td>
<td>The results of initial screening tests are not usually 100% certain, and are often called presumptive results. The results of screening tests are NOT confirmed results. They are often confirmed later, with further diagnostic tests</td>
</tr>
<tr>
<td>Congenital</td>
<td>Present at or shortly after birth.</td>
</tr>
<tr>
<td>Coverage</td>
<td>This is the proportion of people in the eligible group who actually undergo the screening.</td>
</tr>
<tr>
<td>Cut-off level</td>
<td>Screening tests divide people into a group at lower risk of the condition being screened for, and a group at higher risk who are then offered further investigations. Cut off level is a point defined by the programme used to distinguish higher and lower risk.</td>
</tr>
<tr>
<td>Detection rate</td>
<td>Proportion of affected individuals, or carriers who may have a genetic reproductive risk, with positive screening results</td>
</tr>
<tr>
<td>Diagnostic test</td>
<td>Refers to the analytical process involved in obtaining a result. For example the diagnostic test on an amniocentesis sample (invasive procedure) is the karyotype or PCR.</td>
</tr>
<tr>
<td>Disability (WHO definition)</td>
<td>Consequence of impairment in terms of functional performance (i.e. disturbance at the level of the person)</td>
</tr>
<tr>
<td>Disorder</td>
<td>Several words are used to describe illnesses. They are sometimes called diseases, disorders or conditions.</td>
</tr>
<tr>
<td>Dominant inheritance</td>
<td>Every cell contains two copies of each gene. If only one of these copies is altered and one is not, but the person is affected by a characteristic or disorder caused that alteration, the alteration is dominant. A characteristic or disorder caused by a dominant gene alteration only requires one of the genes to be altered for the person to be affected.</td>
</tr>
<tr>
<td>Effectiveness</td>
<td>The extent to which intervention results in the desired outcomes under everyday conditions.</td>
</tr>
<tr>
<td>Embryo</td>
<td>A fertilised ovum (egg) in the early stage of development. In humans the term is reserved for the first eight weeks of development.</td>
</tr>
<tr>
<td>Epidemiology</td>
<td>The study of the distribution of and influences on health-related states and events in populations and the control of health problems, the study of epidemic disease.</td>
</tr>
<tr>
<td>False-negative result</td>
<td>Screening tests divide people into lower and higher risk groups. Some people with a negative screening test result do actually have the condition being screened for. These people are said to have a ‘false-negative’ result.</td>
</tr>
<tr>
<td>False-positive result</td>
<td>Screening tests divide people into lower and higher risk groups. Some people with a positive screening test result do not actually have the condition being screened for. These people are said to have a ‘false-positive’ result.</td>
</tr>
<tr>
<td>Family history</td>
<td>History of a condition in at least one of the following family members: parent, sibling, grandparent, great-grandparent, aunt, uncle, nephew, niece or cousin or child.</td>
</tr>
<tr>
<td>Family origins</td>
<td>A term used to describe a person’s ancestry</td>
</tr>
<tr>
<td>Fetus</td>
<td>In humans, the unborn child after the end of the eighth week of pregnancy to the moment of birth.</td>
</tr>
<tr>
<td>Genetic counselling</td>
<td>Information and support provided by an appropriately</td>
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</tbody>
</table>
trained health professional, to individuals who have known conditions in their families or who are concerned about the future possibility of genetically inherited conditions.

Genetic counsellor
A health professional with specialised training in genetics and counselling who can provide information and support for individuals or families with concerns about a genetic disorder that may run in the family.

Genetic disorders
Conditions that result from alterations in an individual’s genetic makeup. They may be the consequences of alterations in single genes, or in whole chromosomes, parts of which may be lost, duplicated, misplaced or replaced; or may result from the interaction of multiple genes and external factors such as the environment.

Genetic testing
Examination of an individual’s genetic material to identify alterations that may cause a disorder.

Genetics
1. The study of the structure and function of genes.
2. The genetic features which occur in individuals, families and populations.

Gestational age
The duration of an ongoing or completed pregnancy, measured from the first day of the last menstrual period (usually about two weeks longer than that measured from conception). Gestational age is usually measured in completed weeks.

Haemoglobin
The main protein in red blood cells, which carries oxygen from the lungs to the tissues. It consists of four protein subunits called globin chains, each carrying one molecule of haem, which binds and releases oxygen. Haem is red, and this is why blood is red. There are several types of human haemoglobin. Adults have mainly haemoglobin A, which consists of two alpha and two beta globin chains (a2b2), and fetuses have mainly haemoglobin F, which consists of two alpha and two gamma globin chains (a2g2).

Haemoglobin disorders
Mild or serious disorders that can occur in people who have inherited two haemoglobin gene variants. The most common haemoglobin disorders are sickle cell disorders and thalassaemia disorders, also called haemoglobinopathies.

Impairment (WHO definition)
Dysfunction resulting from pathological changes in a system.

Incidence
The number of new instances of a specific condition occurring during a certain period in a specified population.

Inheritance
The passing of familial characteristics from one generation to the next.

Inherited
Having a hereditary characteristic; there are many inherited characteristics, including eye colour, hair colour and health disorders.

Invasive procedure
Invasive procedure - is a method used to obtain a sample, usually to aid diagnosis e.g. amniocentesis and chronic villi sampling are invasive procedures.

Invited group
Those to whom a screening test is offered.

In vitro fertilisation (IVF)
The process whereby an egg is fertilised with sperm in the test tube and then transplanted into a woman’s uterus.

Karyotype
A photomicrograph of an individual’s chromosomes arranged in a standard format showing the number, size, and shape of each chromosome type; used to correlate
chromosomal anomalies with the characteristics of specific diseases. Karyotyping is often used for prenatal diagnosis of conditions such as Down’s syndrome.

**Marker**

An identifiable physical location on a chromosome whose inheritance can be monitored. Markers can be expressed regions of DNA (genes) or some segment of DNA with no known coding function but whose pattern of inheritance can be determined.

**Miscarriage**

Loss of a fetus before the 24th week of pregnancy.

**Morbidity**

The extent of being affected by a disease or condition. In epidemiology, the morbidity rate is the prevalence of a disease within a particular number of the population.

**Morbidity rate**

In epidemiology, the prevalence of a disease within a population, usually expressed as cases per 100,000.

**Mortality/mortality rate**

The incidence of death in a population in a given period.

**Non-invasive**

A procedure that does not require incision into the body or the removal of tissue.

**Placenta**

The structure that provides the fetus with nourishment during development. It is attached to the wall of the uterus and connects to the fetus through the umbilical cord.

**Prevalence**

The proportion of people in a population who have a given disease or attribute.

**Prevalence rate**

The number of people with the condition or attribute, divided by the population at risk.

**Prognosis**

Predicted course and outcome of a disorder, based on all the knowledge related to a specific case, eg, age, sex, the course of the disorder in other patients.

**Quality assurance**

A system for monitoring and maintaining high standards in every aspect of a screening programme.

**Recessive**

Every cell contains two copies of each gene. Each gene contains the information to produce a particular gene product, such as a protein. If a gene is altered, it may no longer code for the gene product. Where an individual has one altered gene copy, the cell will only produce half the usual gene product and may also produce half of the altered gene product. If this does not result in any disorder for the individual, the alteration is described as being ‘recessive’ to the unaltered copy of the gene. An individual with this genetic constitution is said to be a ‘carrier’ of a recessive gene alteration. For a recessive gene alteration to result in a particular characteristic or disorder, both copies of the genes must be altered.

**Risk**

Risk is usually taken to mean the chance of an event happening. It can be expressed in a number of ways.

**Screening**

Screening is a process of identifying apparently healthy people who may be at increased risk of a disease or condition. They can then be offered information, further tests and appropriate treatment to reduce their risk and/or any complications arising from the disease or condition.

**Screening programme**

The whole system of activities needed to deliver high quality screening. It ranges from identifying and informing those to be offered screening through to the treatment and follow up of those found to have abnormality, and support for those who develop disease despite screening.

**Screening test**

A test or inquiry used on people who do not have or have not recognised the signs or symptoms of the condition.
being tested for. It divides people into low and higher risk groups.

**Sensitivity**
This is a measure of test performance. High sensitivity means that the test ‘catches’ as many people with the condition as possible. It is measured as the proportion of those with the condition, who have a positive test result. It is the same as the detection rate.

**Sickle cell anaemia (SS)**
A sickle cell disorder caused by inheritance of two genes for haemoglobin S, which often results in significant health problems and requires treatment. Some affected children can be ‘cured’ by bone marrow transplantation.

**Sickle cell disorders**
A group of inherited disorders that are characterised by sickling of red blood cells when there is a shortage of oxygen. The most common sickle cell disorders are sickle cell anaemia (SS), haemoglobin SC disorder, and haemoglobin S/beta thalassaemia. Sickle cell disorders can cause episodes of acute pain (crisis), anaemia, increased risk of infections, and chest problems. They can be life-threatening, particularly for young children.

**Specificity**
This is a measure of test performance. High specificity means the test has as few false positives as possible. It is measured as the proportion of those without the condition, who have a negative test result.

**Surveillance**
Ongoing observation of the health of individuals or populations.

**Syndrome**
Combination of symptoms and signs grouped together to form a disorder.

**Termination of pregnancy**
The medical expulsion or extraction from the uterus of a fetus in the first, second or third trimester of pregnancy.

**Thalassaemia major**
A group of inherited conditions caused by a reduction in the amount of haemoglobin produced. People with a thalassaemia condition have various degrees of severe anaemia.

**Thalassaemia carrier (also called thalassaemia trait)**
The presence of one thalassaemia gene and one normal haemoglobin gene in an individual. This causes the red cells to be small and individuals may have very mild anaemia, but this does not usually cause any problems.

**True-negative result**
Screening tests divide people into low and higher risk groups. Most of the people with a negative screening test result do not have the condition being screened for. These people are said to have a ‘true-negative’ result.

**True-positive result**
Screening tests divide people into low and higher risk groups. Some people with a positive screening test result do have the condition being screened for. These people are said to have a ‘true-positive’ result.

**Twins**
May be genetically identical (monozygous) when they arise from a single fertilised egg or non-identical (dizygous) when they arise from two separate eggs.

**Uptake**
Is the proportion of people, who when offered a test, take it up.

**Variant**
A change for example in a gene or protein. For example, a variant in a haemoglobin gene resulting in a variant in the haemoglobin the body produces thus causing a sickle cell disease.